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彗星实验技术及其应用

李岗1、吴声敢12,3,4、蔡磊明1,2,3,4,*

1. 浙江省农业科学院农产品质量标准研究所,杭州 310021

2. 浙江省农业科学院 农业部农药检测重点实验室,杭州 310021

3. 浙江省农业科学院 浙江省农药残留检测与控制研究重点实验室,杭州 310021

4. 浙江省农业科学院 浙江省植物有害生物防控省部共建国家重点实验室培育基地,杭州 310021

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摘要: 彗星实验是瑞典科学家 Ostling 和 Johanson 于 1984 年发明的检测毒物 DNA 损伤效应的方法。它经历了从最初的微电 泳技术、中性彗星实验、碱性彗星实验、酶切彗星实验和双向垂直彗尾彗星实验等不断完善的发展过程。在毒理学、遗传学和 环境生态科学等领域有着重要的应用,是经济合作与发展组织(OECD)和欧洲食品安全局等国际组织推荐的测定遗传毒性的 方法之一。彗星实验的关键点包括单细胞悬液的制备、细胞裂解液的成分与比例,低熔点琼脂糖凝胶的浓度,电泳条件等。 在典型应用领域,如蚯蚓、鱼、两栖动物、鼠和人的彗星实验很难找到标准实验方案。成功的彗星实验还需关注,实验设计时 必须包括阳性对照,结果表述时必须有图为证,实验方案可能因物种或细胞而异。

关键词: 彗星实验; 单细胞凝胶电泳; DNA 损伤; 遗传毒性

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The Comet Assay and Its Applications

Li Gang¹, Wu Shenggan^{1,2,3,4}, Cai Leiming^{1,2,3,4,*}

1. Institute of Quality and Standard for Agro-Products, Zhejiang Academy of Agricultural Sciences, Hangzhou 31002, China

2. Agricultural Ministry Key Laboratory for Pesticide Residue Detection, Zhejiang Academy of Agricultural Sciences, Hangzhou 31002, China

3. Key Laboratory for Zhejiang Pesticide Residue Detection and Control, Zhejiang Academy of Agricultural Sciences, Hangzhou 31002, China

4. State Key Laboratory Breeding Base for Zhejiang Sustainable Pest and Disease Control, Zhejiang Academy of Agricultural Sciences, Hangzhou 31002, China

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Abstract: In 1984, the Sweden scientists, O. Ostling and K.J. Johanson, devised the "comet assay" to identify substances that cause DNA damage. Since then, it has experienced refinements that include several improvements such as microelectrophoresis, the neutral comet assay, the alkaline comet assay, the enzyme-modified comet assay, and the two-dimensional perpendicular tail comet assay. This general method is applicable to wide fields of interests

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作者简介:李岗(1973-),男,博士,研究方向为农药毒理学,E-mail: lgxjren@qq.com;

^{*} 通讯作者(Corresponding author), E-mail: hzcaileiming@foxmail.com

including toxicology, genetics, environmental sciences & ecology, and others. Organization for Economic Co-operation and Development (OECD) and the European Food Safety Authority recommend comet assay as an integral part of their genotoxicity testing strategy. The critical procedures in the comet assay involve the preparation of a single-cell suspension, modifications to the lysis buffer, the percentage of low melting point agarose, electrophoresis conditions, etc. It is difficult to standardize protocols for earthworm, piscine, amphibians, murine, and human subjects. Successful comet assay must also include key points such as inclusions of positive controls in experimental design, good comet image as supporting evidence, and the development of special protocols depending on species and cell types.

Keywords: comet assay; single cell gel electrophoresis; DNA damage; genotoxicity

1 发明与发展(Devised and developing comet assay)

彗星实验(comet assay)也叫单细胞凝胶电泳实验(single cell gel electrophoresis, SCGE),是瑞典科学家O. Ostling和K.J. Johanson于1984年发明,刊登在期刊《生物化学与生物物理研究通讯》上,当初叫"微电泳技术(microelectrophoresis)"。实验中,把鼠淋巴细胞L5178Y-S和中国仓鼠的成纤维细胞C1-1悬浮,进行物理辐照后,包埋在琼脂糖胶中,在中性缓冲液中裂解细胞,接着在5V·cm⁻¹电场中电泳5min,再用吖啶橙染色,最后用荧光显微镜镜检。发现DNA向阳极迁移,辐照后的细胞更明显,还发现受辐照细胞能在1h内修复超过50%的损伤^[1]。

1988 年彗星实验迎来第一次重大改进。Singh 进一步发明了低水平损伤 DNA 的定量检测技术, 把人白细胞暴露于 X 射线和 H₂O₂ 后,琼脂糖胶包 埋,接着裂解和电泳都改为碱性条件下进行,因为中 性细胞裂解液和中性电泳液不能检测单链断裂类型 的 DNA 损伤。改进后发现,H₂O₂ 处理的 DNA 迁 移模式比 X 射线的要整齐和同质(homogeneous)。 不同类型的细胞对 DNA 损伤的修复能力有很大差 异。Singh 改进后的彗星实验技术适用检测单个细 胞 DNA 损伤和修复,所需细胞少,1 000 个就足够, 细胞不需要放射性标记,因此,此法适用于任何有核 细胞。还能检测同一种群的细胞对 DNA 损伤物的 不同反应^[2],此后,1990 改名为彗星实验^[3]。

1993 年彗星实验又一次改进。1993 年以前,彗 星实验只能检测 DNA 是否有断裂损伤,不能分辨 非断裂损伤,不能确定 DNA 损伤类型。改进后的 彗星实验,可以确定氧化型非断裂损伤。即解旋后 用核酸内切酶 Ⅲ (thymine glycol DNA glycosylase-Endo III, endonuclease III)处理 DNA,可识别氧化损 伤的嘧啶^[4-6]。同样,用甲酰氨嘧啶 DNA 糖基化酶

fpg (formamidopyrimidine DNA glycosylase, fpg)处理 后,可识别氧化损伤的嘌呤^[7],如8-氧鸟嘌呤,这些 酶可造成氧化损伤部位的 DNA 断裂,增加慧尾的 含量。因此,可用酶切慧星实验检测非断裂型 DNA 氧化损伤,如用 Ro19-8022 和可见光(主要产生 8-氧 鸟嘌呤)暴露肺癌细胞系 A549,在细胞裂解后,用 fpg 处理类核体 30 min 或 45 min,可显著增加慧尾 的浓度^[8-9]。fpg修饰的彗星实验检测大鼠的不同组 织细胞暴露于甲基磺酸甲酯(methyl methanesulfonate, MMS)后,产生高水平的氧化的或烷基化的碱基 或 fpg 伤口^[10]。抗氧化剂对氧化损伤 DNA 的影响 实验表明,非吸烟健康人连续12周暴露在不同的类 胡萝卜素单体下,其淋巴细胞的内源性氧化损伤并 没有受到显著影响,未见高水平的 DNA 断裂损 伤^[11],混合维生素 C、维生素 E 和 B 胡萝卜素干预非 吸烟人和吸烟人,可显著降低二者淋巴细胞的内源 性 DNA 氧化损伤^[12], 而吸烟人的高水平的氧化碱 基在 20 d 后消失了,发现抗氧化剂的种类和剂型对 氧化损伤的 DNA 修复有潜在的影响,如维生素 C 的缓释剂型和快释剂型对 DNA 损伤修复的效果不 同,男性吸烟者只摄入缓释型维生素 C 可以显著减 少 fpg 或核酸内切酶Ⅲ识别的损伤位点^[13]。改进后 的酶切彗星实验还能提供基因的遗传多态性相关的 遗传背景与环境因子信息,如着色性干皮病(xeroderma pigmentosum group D, XPD)基因的编码区 312-751,纯合野生型基因型的彗尾密度比癌症病人 低2倍,暗示维持正常的 DNA 修复活性能抑制癌 症的发生和发展^[14]。

彗星实验常和微核实验同时进行,从不同角度 探索毒物的损伤效应。彗星实验主要测定原始的 DNA 断裂损伤,而微核实验测定的是 DNA 不可修 复的损伤,这些损伤是非修复的或不正确修复的损 伤。2 种实验的结果常呈正相关。例如,非洲爪蟾 幼蛙暴露在甲基磺酸乙酯(ethyl methanesulfonate, EMS)后造成其红细胞 DNA 损伤和微核率增加^[15]。 彗星实验还和其他指标如活性氧(ROS)结合,探索 毒性机制,如乙草胺通过诱导 ROS 造成花背蟾蜍 (*Strauchbufo raddei*)的 DNA 损伤,同时发现 N-乙酰 半胱氨酸(*N*-acetyl-1-cysteine, NAC)和褪黑素(melatonin, MEL)可减轻乙草胺造成的 DNA 损伤^[16]。

自从 Singh 提出碱性彗星实验后,在遗传毒性 检测中得到快速发展。1999年,欧洲国家成立了一 个彗星实验专家组,一致推荐 Singh 的碱性彗星实 验作为检测化学品遗传毒性的标准方法,该法经优 化后,可用于检测单链断裂的 DNA 损伤、碱性不稳 定位点、DNA-DNA 或 DNA-蛋白质交联损伤和单 链断裂的不完全切除修复位点。其特点是:可检测 低水平的 DNA 损伤,只需少量细胞,灵活经济,简 便快速,例如检测 DNA 损伤的修复能力。DNA 损 伤的修复类型有(1)切除修复。包括切除碱基缺失 位点、8 氧鸟嘌呤、单链断裂缺口、大体积的加合物 和环丁烷嘧啶二聚体等,切除修复路径主要受翻译 后修饰调控[17]。(2)重组修复。修复的是双链断裂的 缺口和链间交联。(3)错配修复。修复的是碱基的错 配、插入或缺失^[18-19]。细胞应对 DNA 损伤,通过激 活复杂的信号通路来决定细胞的命运,或者促进细 胞死亡,或修复损伤而存活^[20]。尽管细胞存在这些 修复机制,但是 DNA 的损伤依然保持在较高的水 平,至少在健康的人体中是这种情况。DNA 的修复 能力可看作是突变或癌症的有价值的标志物,因为 差的修复能力与高风险的癌症相关,例如,头颈鳞状 癌细胞的 DNA 修复能力就很差^[21]。DNA 的修复潜 能经常用 DNA 芯片或 RT-PCR 方法,测定修复通路 基因的转录水平,但是它的酶活性不仅依赖转录或 翻译水平,还要依赖表型分析的结果^[2]。检测修复 过程动态的最简单的方法是进行彗星实验,暴露于 DNA 损伤物后,在不同的时间点取样测定,进行挑 战实验(challenge assay)。例如,用水溶性的 ß 胡萝 卜素或番茄红素培养的淋巴细胞,暴露在博来霉素 (bleomycin)或 H₂O₂下,结果表明 ß 胡萝卜素能保护 DNA 链,防止其断裂,但是不能阻止其碱基的氧化 损伤^[23]。而 β 隐黄素(β-cryptoxanthin)能保护 HeLa 细胞和 Caco-2 细胞免受 H₂O₂ 诱导的 DNA 损伤, 呈现明显的 DNA 修复效应。因此,标准的彗星实 验可用来测定细胞对 DNA 损伤的修复能力。

2000年,欧洲食品安全局主张采用系列实验来

评估化学品的遗传毒性,即首先对化学品进行细菌 回复突变实验(后采用 OECD TG471 准则^[24])和体外 微核实验(后采用 OECD 487^[25]),只要有一个实验结 果呈阳性,再进行 3 个体内实验,包括哺乳动物红细 胞微核实验(即 OECD TG474 准则^[26])、彗星实验(即 OECD TG489 准则^[27])和转基因啮齿动物细胞基因 突变实验(即 OECD TG488 准则^[28])。

2014年,经济合作与发展组织(OECD)专门发 布哺乳动物体内彗星实验准则即 OECD TG489^[27], 内容包括实验原理、局限性、对照信息、方法的详细 描述和彗星图象定量打分(推荐采用的自动和半自 动软件)。其中重要的是对细胞彗星图象进行定义, 分成3类,可打分的(scorable),非打分的(nonscorable)和刺猬形(hedgehog)。可打分的彗星细胞 定义为慧核和慧尾清晰且不与其他细胞重叠的细 胞。彗星评价采用慧尾 DNA 的百分数或慧尾强度 (tail intensity, TI),表示 DNA 的断裂量,再结合校正 曲线和已知条件即每个 Gy 可诱导每 10° Dalton DNA 中0.3 个断裂,可计算实际 DNA 的断裂频率, 校正曲线是 gamma 射线或 X-ray 的剂量(0~10 Gy) 为自变量与慧尾 DNA 的百分数为因变量之间建立 的线性关系^[29-30]。刺猬形细胞 DNA 也叫鬼魂细胞 (ghost cell)或云状细胞,认为它是一种严重损伤的细 胞(慧核小或不存在,大的弥散状尾巴)。通过图像 分析得到的 TI 不可靠,对刺猬细胞应该单独评价, 它的形成机制还不清楚。大鼠肝细胞彗星实验表 明,它可能是破碎细胞时的机械损伤或试剂诱导的 细胞毒性[31]。刺猬形彗星也不能认为是凋亡细胞或 细胞毒性。这样的慧星常表示连续的 DNA 损伤, 应该对其进行全面评估[32]。

2 技术要点(Methodical critical steps)

彗星实验经过不断改进,其共同基本步骤包括 制备悬浮细胞、凝胶包埋、细胞裂解、解旋、电泳、中 和、染色和图象分析。体外彗星实验,先获得悬浮细 胞后暴露处理。体内彗星实验,制备悬浮细胞前实 验个体进行暴露处理。其中实验成败的关键步骤应 包括:

2.1 制备悬浮细胞

获得悬浮细胞因物种而异。人、鼠、鱼等常从其 循环系统中获取细胞,如血细胞或淋巴细胞。

蚯蚓的小细胞常用于彗星实验,小细胞来自蚯 蚓的体腔液,是体腔液中含量最丰富的细胞,类似于 脊椎动物的白细胞。蚯蚓体腔细胞的收集方法,包 括提取液法、穿刺法、电击法和超声波法等^[27,33]。其 中提取液法最常用,其包含:5 mmol·L⁻¹ Na,-EDTA (乙二胺四乙酸)或 EGTA(乙二醇二乙醚二胺四乙 酸)、50.4 mmol·L⁻¹或10 mg·mL⁻¹愈创木酚甘油醚、 95% 生理盐水(即 0.85 g NaCl 溶于每 100 mL H₂O) 或磷酸盐缓冲液(PBS)和5%乙醇或甲醇,pH7.3~ 7.5。其中 EDTA 或 EGTA 和生理盐水或 PBS 的作 用是维持活细胞正常活性。愈创木酚甘油醚的作用 在于溶解细胞粘液,释放细胞。乙醇或甲醇的作用 刺激蚯蚓释放体腔液。

植物彗星实验最大困难是获得悬浮细胞或类 核,常用根和叶进行^[34-35],实验方案因组织或物种而 异^[36],通常的碱性溶液提取方法不能从植物细胞中 释放完整的 DNA,用机械破碎或化学处理的方法可 对 DNA 造成新的损伤,叶片中的高浓度色素如叶 绿素和代谢物对分离 DNA 可能造成新的损伤,所 以,弃用叶片细胞而选用根尖细胞,但是高频率分裂 的细胞对实验又产生影响。尽管植物细胞彗星实验 比较困难,但是也有成功的例子,例如Cr(VI)对豌豆 (Pisum sativum) DNA 损伤实验^[37], Cd-Zn 对烟草的 影响[38]。

目前认为悬浮细胞中的活细胞应不少于70% ~75%,才能得到可靠的结果[39],通常用苔盼蓝染色 法检测活细胞百分数。

2.2 凝胶包埋

常用低熔点琼脂糖凝胶包埋细胞,在三层胶法 中,还用正常熔点凝胶固定低熔点胶,防止在后继的 浸泡步骤中胶从载胶板上脱落。所以,低熔点胶的 浓度至关重要。不同人外周淋巴细胞和淋巴母细胞 系 TK-6 的结果表明,胶浓度在 0.6%~0.8% 比较合 适,太低,胶不稳定,太高,损害慧尾。胶浓度与慧尾 长度呈明显的线性关系。损伤细胞对胶浓度更敏 感。胶不能反复熔化,应该分装保存^[40]。OECD 建 议低熔点琼脂糖凝胶为 0.5% ~1%, 不建议低于 0.45% (OECD 489)^[27]

2.3 细胞裂解

对包埋在低熔点胶中的完整细胞进行裂解,常 用细胞裂解液的通用配方是:10 mmol·L⁻¹ Tris-HCl, 100 mmol·L⁻¹ Na₂EDTA, 2.5 mol·L⁻¹ NaCl, 10% \equiv 甲基亚砜(DMSO),1% Triton X-100, pH 10,裂解条 件是4℃、至少60 min。其中 Tris-HCl 是缓冲体系, 维持稳定的 pH 值。Na, EDTA 是变性剂和稳定剂。 DMSO 是 DNA 保护剂,防止自由基对 DNA 的损 伤。Triton X-100 是表面活性剂,破坏细胞,类似的 有 SDS(十二烷基硫酸钠,一种强变性剂和蛋白溶解 剂)和肌氨酸钠(sodium carcosinate),也有在裂解液中 加入蛋白酶 K 和氯化钙, 如提高对口哨蛙(Eleutherodactylus johnstonei)细胞的裂解能力^[41-42]。

| Table 1 Positive c | ontrol substances and its target tis | sues in comet a | assay recommended by OECD 489 |
|--------------------|--|-----------------|--|
| 中文 | 英文 | CAS | 靶组织 |
| Chinese name | English name | CAS number | Target tissues |
| 甲基磺酸乙酯 | Ethyl methanesulfonate, EMS | RN 62-50-0 | 全部组织 Any tissue |
| 乙基亚硝基脲 | Ethyl nitrosourea | RN 759-73-9 | 肝 Liver、胃 Stomach、 十二指肠 Duodenum、空肠 Jejunum |
| 甲基磺酸甲酯 | Methyl methanesulfonate, MMS | RN 66-27-3 | 肝 Liver、胃 Stomach、十二指肠 Duodenum、 空肠 Jejunum、肺 Lung、 支气管肺泡细胞 Bronchoalveolar lavage cells、 肾 Kidney、膀胱 Bladder、睾丸 Testis、 骨髓 Bone marrow、血液 Blood |
| N-甲基-N' -硝基-N-亚硝基胍 | N-Methyl-N' -nitro-N-nitrosoguanidine, MNNG | RN: 70-25-7 | 胃 Stomach、十二指肠 Duodenum、空肠 Jejunum |
| 1,2-二甲基肼二盐酸盐 | 1,2-Dimethylhydrazine 2HCl | RN 306-37-6 | 肝 Liver、小肠 Intestine |
| N-甲基-N-亚硝基脲 | N-methyl-N-nitrosourea, NMU | RN 684-93-5 | 肝 Liver、骨髓 Bone marrow、血液 Blood、 肾 Kidney、胃 Stomach、空肠 Jejunum、脑 Brain |

表1 OECD 489 推荐的彗星实验阳性对照及其靶组织

注: OECD 489 为经济合作与发展组织在 2014 年 9 月 26 日发布的化学品试验导则:哺乳动物体内碱性彗星试验。

Note: OECD 489 has been published by OECD on 26th September 2014. It is a guideline for the testing of chemicals: in vivo mammalian alkaline comet assay

| | | | Table 2 | Summary of comet assay par | rameters | s on amp | hibians | | | | | |
|--|--|-----------------------------|-------------------------|---|----------|----------------------------|----------------|-------------------|-------------------------|--|-------------------------|------------|
| 实验生物 Experimental | 毒物 Contaminants | 细胞 Cell | 胶浓度/% | 裂解液 Lysis buffer | | 裂解条件 ysis conditi | suc | 解放 Unw conc | 终件 inding litions | 电泳务 Electrofc conditi | 针件 oresis ons | 参考文献 |
| model | tested | type | Agarose/% | composition | Hd | 温度/℃ Temp./℃ | 时间 Duration | pH Du | 时间/min tration/min | 电压 Voltage | 时间 Duration | vererences |
| 绿蛙 Lithobates clamitans 美洲蟾蜍 Anaxyrus americanu 美洲牛蛙 Lithobates catesbeianus (tadpoles | MMS 阿特拉津 Atrazine 草甘膦 Glyphosate 嗪草酮 Metribuzin | 血液 Blood | 0.4 0.5 0.5 | 2.5 mol·L⁻¹ NaCl, 100 mmol·L⁻¹ Na₂ EDTA, 10 mmol·L⁻¹ Tris, 10% DMSO, 1% Na- sarcosinate | 10 t | Room emperature (RT) | 2 h | 13 | 15 | 25 V (265 ~ 270 mA) | 20 min (37 °C) | [96-98] |
| 美洲蟾蜍 Anaxyrus americanu: 绿畦 上ithobates clamitans 北方豹蛙 Lithobates pipiens | s 多种 Several pollutants | 血液 Blood | 0.4 | 2.5 mol·L⁻¹ NaCl, 100 mmol·L⁻¹ Na₂ EDTA, 10 mmol·L⁻¹ Tris, 10% DMSO, 1% Na- sarcosinate, 1% Triton X-100 | 10 | RT | 2 h | 13 | 15 | 25 V (265 ~ 270 mA) | 20 min (37 °C) | [57] |
| 六趾蛙 Euphlyctis hexadactylus | 含硫溴料 Sulfonated dyes | 血液 Blood | 0.75 | 2.5 mol·L ⁻¹ NaCl, 100 mmol·L ⁻¹ Na ₂ EDTA, 10 mmol·L ⁻¹ Tris, 10% DMSO, 1% Triton X-100 | 10 | RT | 2 h | 13 | 25 | 20 V (300 mA) | 20 min | [48] |
| 非洲爪蟾 Xenopus laevis | 电磁辐射 High peak-power pulsed electromagnetic field | 血液 Blood | 0.5 | 2.5 mol·L ⁻¹ NaCl, 10 mmol·L ⁻¹ Tris, 1% Triton X-100, 100 mmol·L ⁻¹ Na ₂ EDTA | 10 | 22 | 25 min | 13 | 20 | 1.9 V·cm ⁻¹ | 20 min (4 °C) | [59] |
| 黑我侧褶蛙 Pelophylax nigromaculatus | 樹 Lead | 睾丸细胞 Testicular cells | 03 | 2.5 mol·L ⁻¹ NaCl, 10 mmol·L ⁻¹ Na ₂ EDTA, 10 mmol·L ⁻¹ Tris, 10% DMSO, 1% SDS, 1% Triton X-100, pH 10 | 10 | 4 | 2 ~4 h | 10 | 30 (4 °C.) | 22 V (220 mA) | 30 min (4 °C) | [66] |
| 温室蛙 Eleutherodactylus johnstonei | 博来霉素、 4-硝基-1-氧-喹啉、草甘膦 Bleomycin, 4-nitro-quinoline-1-oxide, Glyphosate | 血液 Blood | <i>L</i> .0 | 50 mmol·L ⁻¹ Tris, 10 mmol·L ⁻¹ CaCl ₂ , 0.04 g·mL ⁻¹ proteinase K | ∞ | 6±2 | 10 min | 13 | 25 | 22 V (220 mA, 0.9 V·cm ⁻¹) | 30 min (6 °C ±2 °C) | [41-42] |
| 注:Na ₂ EDTA 表示乙 Note: Na ₂ EDTA stand | 二胺四乙酸二钠;DMSO表 Is for edetate disodium; DMS | 示二甲基亚 SO stands for | 砜;SDS 表示 dimethyl su | 六十二烷基硫酸钠。 (foxide; SDS stands for sodium dodecyl s | sulfate. | | | | | | | |

常见两栖动物彗星实验条件

表 2

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2.4 碱性解旋

解旋液和电泳液是同一种溶液,OECD 489 建 议配方和条件是:1 mmol·L⁻¹ Na₂EDTA,300 mmol· L⁻¹ NaOH,pH≥13,4 ℃至少解旋 20 min,电泳条件 是4 ℃,0.7 V·cm⁻¹,20 ~40 min。DNA 迁移与电泳 时间呈线性相关。对人外周淋巴细胞和淋巴母细胞 系 TK-6,以电压 1.15 V·cm⁻¹,电泳 20 min 为佳,低 于 0.49 V·cm⁻¹,慧尾不能展开,大于 1.48 V·cm⁻¹,慧 尾从慧核上断离,人为增加损伤程度^[40],需要记住的 常识是,同样的电泳迁移距离时,单位电压与时间的 乘积是个常数。

2.5 细胞密度

彗星的数量或密度影响图像分析与评价,因 此细胞或彗星不能重叠,以免影响彗星计分。为 便于统计分析,需要对至少50个细胞进行图象分 析。而有效实验能给出定性判断的最少细胞数为 30个。

2.6 阳性对照

彗星实验必须设置阴性对照和阳性对照,保证 实验设计的合理性,实验结果的可靠性和有效性。 OECD 489 推荐的哺乳动物常用阳性对照,包括甲 基磺酸乙酯(EMS)等,见表1。

这些阳性对照,可能也适用于非哺乳动物,如海 鳟(Salmo truta)和北极嘉鱼(Salvelinus alpinus)的精 细胞暴露在甲基磺酸甲酯(MMS)中,可造成 DNA 损伤,并且后代的骨胳发育异常,表明这种损伤具有 遗传性^[43]。三刺鱼(Gasterosteus aculeatus)精子在体 外受精前,体外暴露在 MMS 中,可造成后代的胚胎 发育异常^[44]。另外,4-硝基喹啉 N-氧化物(4-Nitroquinoline N-oxide)是模式遗传效应毒物,会造成斑 马鱼 DNA 损伤和产卵量下降^[45-46]。

成功的彗星实验要考虑3个方面因素,一是细胞的类型如淋巴细胞、生殖细胞或细菌细胞、二是诱导物类型如X射线、伽马射线、乙醇、乙醛、疾病、生活类型因素。三是方法学因素如电泳槽、载胶板、分析参数等。

3 典型应用(Main application field)

用"comet assay"作关键词在 2016 年的 PubMed 和 Web of Science 中搜索,可得近万篇期刊文章。 涉及的主要应用领域包括:毒理学 30%,遗传学 15%,环境生态科学 11% 和应用微生物技术 9%。 彗星实验还有专门的交流网站 http://www.cometassay.com/ $_{\circ}$

3.1 两栖动物彗星实验

绿蛙(Lithobates clamitans)和非洲爪蟾(Xenopus laevis)等的成体或幼体的红细胞常用于体内彗星实验,见表2。

测定的毒物效应包括,除草剂^[42]、杀虫剂^[47]、 MMS^[15]、EMS^[15]、苯并芘^[15]、硫化染料^[48]、金属^[49]、石 油化工产物^[50]、持久性有机污染物(persistent organic pollutants, POPs)^[51]、抗生素^[41]、二甲基亚砜(dimethyl sulfoxide, DMSO)^[41]、污染的湖水^[52]、煤矿^[53]、垃圾 场^[54]、污泥^[55]、污水^[56-57]、城市固体废物的焚烧物^[58] 和电磁场的影响^[59]等。

两栖动物彗星实验方案,差别较大。1997年之后的实验,细胞裂解液中才开始含有洗涤剂(表面活性剂)如 Triton X-100、DMSO、SDS、肌氨酸钠、蛋白酶 K 和氯化钙等,来提高对细胞的裂解能力。细胞裂解时间 25 min 至 7 d。细胞裂解温度是 4 °C,而 2005年之前是室温。低熔点琼脂糖浓度通常为 0.5%,范围通常是 0.4% ~1.0%,浓度越高,慧尾越 $\mathcal{P}^{[60]}$ 。解旋变性通常是 pH>13条件下 5~40 min,时间越长,慧尾越多^[60]。电泳电压是 18~27 V,电流是 300 mA,也有用 V·cm⁻¹表示的。这些实验方案的差异,限制了实验结果的可比性。

3.2 鱼类彗星实验

最早的鱼类彗星实验是用加拿大五大湖的棕鲷 鱼(brown bullhead, Ameiurus nebulosus)和普通鲤鱼 (common carp, Cyprinus carpio)对湖中沉积物的暴露 实验,提取它们的红细胞进行 singh 于 1988 年提出 的碱性彗星实验^[61]。鱼的血细胞是最常用的彗星实 验材料,因为易得,易分离,而且所有鱼的血液细胞 都是有核细胞。而其他组织细胞如肝、肾、腮^[62-64]、 精细胞^[65]也经常用于彗星实验^[65]。鱼的酶切彗星实 验首次运用是在 2003 年^[66],涉及的细胞有:血细胞、 肝细胞、腮细胞^[67]和细胞系^[68]等。

截止 2017 年,已经有 90 多种鱼用于彗星实验, 实验类型有体内实验(*in vivo*)^[69]、体外实验(*in vitro*)^[68]、离体实验(*ex vivo*)^[44]和原位实验(*in site*)^[70]。 涉及的毒物包括 POPs^[71]、金属^[64]、农药^[72]、药物^[73]、 内分泌干扰物如四溴双酚-A^[74]、纳米颗粒^[75]、生物 毒素^[76]、核辐射^[77]和紫外线^[78]等。

鱼类像两栖类一样,也没有标准的彗星实验方 案,见表3。但是,鱼的室内实验和野生鱼的生物监 测实验中,要求使用标准的采样方案^[79]。

| 常见鱼类彗星实验条 |
|-----------|
| 表 3 |

Table 3 Summary of comet assay parameters on fish

| 参考文献 | Kerences | [001] | [101] | [95] | [102] |
|-----------------------------|------------------------|--|--|--|--|
| \$件 oresis ons | 时间 Duration | 25 min (3 °C) | 15 min | 5 ~ 40 min | 30 min |
| 电泳多 Electrofi conditi | 电压 Voltage | 20 V (300 mA) | 20 V (300 mA) | 0.66 V • cm ⁻¹ | 25 V (300 mA) |
| 旋条件 winding nditions | 时间/min Duration/min | 15 | 30 | 5 ~40 | 60 |
| 」 第 10 10 | Hd | ~13 | 12.3 | >13 | $\frac{1}{3}$ |
| suo | 时间/h Duration/h | 0 | - | - | _ |
| 裂解条件 Lysis conditi | 温度/°C Temp./°C | RT | 4 | 4 | -1 4 |
| | Hq | 10 | 10 | 0 | 原文未见 |
| 裂解液 Lysis buffer | composition | 2.5 mol·L ⁻¹ NaCl, 10 mmol·L ⁻¹ Tris-base, 0.1% sodium sarcosinate | 2.5 mol·L⁻¹ NaCl, 100 mmol·L⁻¹ Na₂ EDTA, 10 mmol·L⁻¹ Tris, 1% Triton X-100, 10% DMSO | 2.5 mol·L⁻¹ NaCl, 100 mmol·L⁻¹ Na₂ EDTA, 10 mmol·L⁻¹ Tris, 1% sodium sarcosinate, 1% Triton X-100, 10% DMSO | 2.5 mol·L⁻¹ NaCl, 100 mmol·L⁻¹ EDTA, 10 mmol·L⁻¹ Tris base, 1% sodium lauryl sarcosinate, 1% DMSO 1% DMSO |
| 胶浓度% Agarose | % | 0.75 ey | 0 | 0 5 | 原文未见 mm |
| 细胞 Cell | type | 血、肝、肾 llood, liver, kidr | الله Blood | 血血 Blood | 精子 Spermatozoa |
| 毒物 Contaminants | tested | 黄曲霉毒素 B1 Aflatoxin B1 E | 多环芳烃(PAHs)和金厚 Polycyclic aromatic hydrocarbons (PAHs) and metals | 苯唑卡因 Benzocaine | 双(氮丙啶基) 甲氨基膦硫化物 Bisazir |
| 实验生物 Experimental | model | 虹鳟 Oncorhyncus mykiss 斑点叉尾鮰 Ictalurus punctatus | 蜎 Pholis gunnellus | 尼罗罗非鱼 Oreochromis miloticus | 海七鳃曼 Petromyzon marinus |

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| 製師業 製師業 単業業件 世球条件 世球条件 世球条件 Lysis buffer Lysis conditions Conditions Conditions Electroticesis 参考文 Lysis buffer pH 通販欠 时何和 Electroticesis 参考文 25 mol.1 ⁻¹ Na; 1% Triton X. a 4 1 12 20 Notage Duration 1% Innot L ⁻¹ Tris, 1% Triton X. a 1 12 20 32 V Notage Ionin 1% Innot L ⁻¹ Tris, 1% Triton X. a 1 12 20 32 V Notage Ionin 1% Triton X-100, 13 4 1 12 20 32 V Notage Ionin 1% Triton X-100, 13 4 1 13 32 V Notage Ionin Ionin 1% Triton X-100, 13 4 1 13 32 V Notage Ionin | |
|---|--|
| Composition PH \underline{MMC} \underline{BHMC} \underline{BHMC} \underline{BHMC} \underline{MHM} | 细胞 胶浓度% Cell Agarose |
| 25 mol·L ⁻¹ NaCh, 10 mmol·L ⁻¹ Na, PaCH, 10 mmol·L ⁻¹ Tins, 1% Triton-X, 1% lauroy serosine sodium salt, 4 mmol·L ⁻¹ lithium diiodosalicytate 8 4 1 12 20 25 V (300 mA) 10 min [65] 1% lauroy serosine sodium salt, 4 mmol·L ⁻¹ lithium diiodosalicytate 13 4 1 12 20 300 mA) 10 min [65] 1% lauroy serosine sodium salt, diiodosalicytate 13 4 15 300 mA) 10 min [67] 10% DMSO 13 4 15 310 mA) [310 mA) 10^{31} mB [60] 25 mol·L ⁻¹ NaCl, 10% DMSO 13 310 mA 310 mA) 15 mB 300 mA) 15 mB 10^{31} mB 15^{31} mB 15^{31} mB 10^{31} mB </th <th>type /%</th> | type /% |
| $ \begin{array}{cccccccccccccccccccccccccccccccccccc$ | 精子 原文未见 permatozoa nm |
| $ \begin{array}{cccccccccccccccccccccccccccccccccccc$ | 细胞系 Cell lines |
| 2.5 mol·L ⁻¹ NaCl, 100 mmol·L ⁻¹ EDTA, 10 mmol·L ⁻¹ Tris-HCl, 10 4 1 >13 20 $\frac{25 \text{ V}}{(300 \text{ mA})}$ 20 min [75 10% DMSO, 10% DMSO, 1% Triton X-100 | 血、肝 ood and liver 1.0 |
| | 、颰细胞系 Heart and 0.9 ill cell lines |

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续表3

注: nm 表示原文未提到。 Note: nm stands for not mentioned.

| | | | Table 4 | Summary of comet assay pa | arametei | rs on ear | rthworm | | | | | |
|---|--|----------------------|-------------|---|----------|---------------------|--------------------|-----------------|--------------------------|------------------------------|-------------------|------------|
| 实验生物 Experimental | 毒物 Contaminants | 细胞 Cell | 胶浓度% | 裂解液 Lucic but the commonition | | 裂解条体 ysis condit | tions + | 型 Unw Con | e条件 inding litions | 电泳条 Electrofo conditio | 件 resis ins | 参考文献 |
| model | tested | type | Agalose /0 | nonvodulo tanno svett | Hq | 温度//C Temp.//C | 时间/h Duration/h | Hd | 时间/min Duration/min | 电压 Voltage | 时间 Duration | Vereiences |
| 远盲蚓 Amynthas diffingens 灰蚓 Aporrectodea caliginosa 草间丛林蚓 bendrodrilus rubidus 赤子愛胜蚓 Eisenia fetida | 縮 Cadmium | 体腔细胞 Coelomocytes | Q Q | 2.5 mol·L ⁻¹ NaCl, 100 mmol·L ⁻¹ EDTA, 10 mmol·L ⁻¹ Tris, 10% DMSO, 0.2 mol·L ⁻¹ NaOH, 1% Triton X-100 | 10 | 4 | <u>5</u> | 13 | 5 | 35 V (300 mA) | LO min | [8] |
| 灰鵐 Aporrectodea caliginosa | 锏、镉、五氯酚 Copper, cadmium, and pentachlorophenol | 体腔细胞 Coelomocytes | 0.8 | 2.5 mol·L ⁻¹ NaCl, 100 mmol·L ⁻¹ EDTA, 10 mmol·L ⁻¹ Tris-HCl, 10% DMSO, 1% Triton X-100 | 10 | 4 | - | 13 | 15 | 35 V (300 mA) | 20 min (4 °C.) | [82] |
| 安德爱胜蚓 <i>Eisenia</i> andrei Bouché | 大气颗粒物、 PAHs、Polluted air containing PAHs | 体腔细胞 Coelomocytes | 1.0 | 2.5 mol·L ⁻¹ NaCl, 100 mmol·L ⁻¹ EDTA, 10 mmol·L ⁻¹ Tris, 10% DMSO, 1% Triton X-100, 0.2 mol·L ⁻¹ NaOH | 10 | 4 | 过夜 Overnight | 13 | 25 | 0.74 V •cm ⁻¹ | 30 min (RT) | [104] |
| 赤子爱胜蚓 Eisenia fetida | 叱虫 味 Imidacloprid | 体腔细胞 Coelomocytes | <i>L</i> .0 | 2.5 mol·L⁻¹ NaCl, 10 mmol·L⁻¹ Tris, 100 mmol·L⁻¹ Na₂ EDTA, 1% Na-sarcosinate, 10% DMSO, and 1% Triton X-100 | 10 | 4 | - | 13 | 30 | 25 V (300 mA) | 15 min (RT) | [105] |
| 赤子爱胜蚓 Eisenia fètida | 桉树挥发性有机物 Volatile organic compounds from Eucalyptus grandis | 体腔细胞 Coelomocytes | 原文未见 nm | 2.5 mol·L⁻¹ NaCl, 10 mmol·L⁻¹ Tris, 100 mmol·L⁻¹ Na₂ EDTA, 1% Na-sarcosinate, 10% DMSO, and 1% Triton X-100 | 10 | 4 | - | >13 | 30 | 25 V (300 mA) | 30 min (RT) | [80] |

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表4 常见蚯蚓彗星实验条件

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| | | | Tat | whee 5 Summary of comet a | tssay par | umeters or | ı human | | | | | |
|-------------|----------------------------|--|----------|---|----------------------|------------|-----------|--------|---|---------------------------|------------------------|------------|
| | | | | | | 刻艇? | 各体 | [期] | 旋条件 | 电泳务 | \$件 | |
| 实验生物 | 毒物 | 细胞 | 胶浓度/ | ?//6 裂解液 | | Lysis cor | ditions | Un | vinding | Electrof | Dresis | 参考文献 |
| rxpermentat | Contaminants test | ed Cell type | Agarose | e/% Lysis buffer composition | [d | H Temn/ | C 时间/h | Hq | 回 [1] [1] [1] [1] [1] [1] [1] [1] [1] [1] | unun 电压 in Voltage | uus 时间 Duration | keferences |
| | | 11111111111111111111111111111111111111 | TTT. | 2.5 mol·L ⁻¹ NaCl, | | | | | | | | |
| \prec | 姜黄素 | 孔大小中小店 脑细胞系 | 承 053 | 100 mmol·L ⁻¹ EDTA, | - | V | - | 1 | 30 | 300 m A | 30 min | [106] |
| Human | Curcumin | RCDA D | | 10 mmol·L ⁻¹ Tris, pH 10 | - | • | - | - I | 0 | | | [001] |
| | | NING | | and 10% DMSO with 1% Trito | n X-100 | | | | | | | |
| | | | | 2.5 mol·L ⁻¹ NaCl, 0.1 mol·L ⁻¹ N | a ₂ EDTA, | | | | | | | |
| 4 | | 肺上皮细胞3 | WZ. | 0.1 mol·L ⁻¹ Tris base, | | | 计游 | | | 1-mo-11 0 0 | | |
| Human | CeO ₂ -NP, KBrO | ³ BEAS-2B | 0.68 | 3 11% Triton X-100, | 1 | 0 4 | Overnight | 13 | 30 | (300 mA) | 20 min (4 °C) | [107] |
| | | BEAS-2B (lun | (gi | 1% lauroyl sarcosinate, | | | | | | | | |
| | | | | 10% DMSO | | | | | | | | |
| | 味精 | 外国血淋口细 | 旧 | 2.5 mol·L ⁻¹ NaCl, 100 mmol·L ⁻ | ¹ EDTA, | | | | | 17 SC | | |
| (1) | Monosodium | ערבייתייד דייייידייייד | 0.65 | 5 10 mmol·L ⁻¹ Tris, | 1 | 0 4 | 1 | >13 | 20 | V C7 | 20 min | [108] |
| плинан | glutamate (MSG |) Lympnocyter | ~ | 10% DMSO, 1% Triton X- | 100 | | | | | | | |
| | | | | 2.5 mol·L ⁻¹ NaCl, | | | | | | | | |
| \prec | 脱氧核糖核酸酶 | 等 精子 | 0.66 | 10 mmol·L ⁻¹ EDTA, | - | 4 | 50 | 13 | 5 C | 20 V | 4 min | [1001] |
| Human | DNAse | Sperm | 00.0 | 10 mmol·L ⁻¹ Tris, 10% DM | SO, | r 0 | 5 | 1 | 1 J | (1.0 V·cm ⁻¹) | F | [701] |
| | | | | 1% Triton X-100 | | | | | | | | |
| 4 | た 坊 | 中省 | | 2.5 mol \cdot L ⁻¹ NaCl, 10 mmol \cdot L ⁻¹ | EDTA, | | | | | 75 V | | |
| Human | Desticides | Blood cells | 0.71 | 10 mmol·L ⁻¹ Tris, 10% DM | SO, 1 | 0 4 | 1 | >13 | 20 | 00 m 0 0 V.C | 20 min | [110] |
| | | | | 1% Triton X-100 | | | | | <u>ر</u> | | (" | |
| | | 外围血细胞(白丝 | 田) | 2.5 mol·L ⁻¹ NaCl, | | | | | | | | |
| \prec | Diesel engine | Leukocytes ir | 1 0.98 | 100 mmol·L ⁻¹ EDTA, | ÷ | 4 | 25 | ⊗13 | 20 | 300 mA, 1.0 V·cı | n ⁻¹ 25 min | [111] |
| Human | exhaust | peripheral bloc | þ | 10 mmol·L ⁻¹ Tris, | | | | | | | | |
| | | | | 10% DMSO, 1% Triton X- | 100 | | | | | | | |
| ~ | た話 | 口胶细胞 | | 2.5 mol·L ⁻¹ NaCl, 100 mmol·L ⁻ | ¹ EDTA, | | | | | 75 V | | |
| Human | Desticides | H JHE FHIJIE Buccal cells | 0.5 | 10 mmol·L ⁻¹ Tris, pH 10 | | 0 4 | 24 | 13 | 20 | (300 mA) | 20 min | [112] |
| THUNK | CONTONIO I | | | 10% DMSO, 1% Triton X- | 100 | | | | | | | |
| | | | | | | | | | | | | |

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表5 常见人彗星实验条件

注:CeO2-NP 表示 CeO2 纳米颗粒。 Note: CeO2-NP stands for CeO2 Nano particles; DNAse stands for deoxyribonuclease; BCPAP stands for a papillary thyroid cancet-derived cell line.

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报 生 态 毒 理 学

| 6 常见鼠彗星实验条件 | v of comet assay parameters on murine |
|-------------|---------------------------------------|
| 表 6 | Summary of |
| | Table 6 |

| 实验生物 | 毒物 | 細胞 | 胶浓度% | 裂解液 | Ę, | 裂解条件 ysis conditior | IS | 解加Unv | 6条件 /inding | 电泳条 Electrofo | 件 resis | 参考文献 |
|-----------------------|------------------------|-----------------------------|-------------|--|----|------------------------|-------------------|---------------|------------------------|---|-------------------------|------------|
| Evnerimental model | Contaminants tasta | Gall type | Agaroca/0/ | Lusis huffer commention | | | | con | ditions | conditio | SU | Beferences |
| | | a con spe | n peo mar | | Hq | 温度/°C Temp./°C D | 时间/h uration/h | PH | 时间/min Duration/min | 电压 Voltage | 时间 Duration | |
| 理上されよ母 | 右雷佐生 | | | 100 mmol·L ⁻¹ EDTA, | | | | | | | | |
| 墙士日化小 厩 | Dexrazoxane | 骨髓细胞 | | 2.5 mol·L ⁻¹ NaCl, | ¢ | - | - | 5 | 0 | $(7 \sim 1.0 \text{ V} \cdot \text{cm}^{-1})$ | 25 min | [[[]]] |
| Mouse | 表阿霉素 | Bone marrow ce | CH-U SII | 10 mmol·L ⁻¹ Trizma base, | 10 | 4 | _ | $\frac{1}{2}$ | 70 | (300 mA) | (4 °C) | [113] |
| (Swiss aldino) | Epirubicin | | | 1% Triton X-100, 10% DMSO | | | | | | | | |
| | SiO ₂ 纳米颗粒、 | 心、肝、肺、 | | 2.5 mol·L ⁻¹ NaCl, | | | | | | | | |
| 小鼠 | Si 纳米颗粒 | 肾、脑 | 0 (6 | 10 mmol·L ⁻¹ Tris, | 0 | - | - | 01 | ¢, | 25 V | | L4 1 1 |
| Mouse (Tuck-Ordinary) | SiO_2 , | Heart, liver, lun | وہ 0.00 | 100 mmol·L ⁻¹ EDTA, | 10 | 4 | 1 | 10 | 70 | (300 mA) | um 02 | [114] |
| | Silica nanoparticles | kidney, brain | | 1% Triton X-100, 10% DMSO | | | | | | | | |
| | | | | 1% Sodium sarcosinate, 2.5 mol·L ⁻¹ NaCl, | | | | | | | | |
| 小鼠 | 15 °C 33 °C | 脾淋巴细胞 | 0.45 | 100 mmol $\cdot L^{-1}$ Na ₂ EDTA, | 0 | - | - | 5 | 00 | 75 17 | | [116] |
| Mouse | 1 77 77 61 | Spleen lymphocy | /te 0.45 | 10 mmol·L ⁻¹ Tris, | 10 | 4 | 1 | <u>دا</u> ~ | 07 | A C7 | um 02 | [611] |
| | | | | 1% Triton X-100 | | | | | | | | |
| 5 - + | 声 沉岫 | nden JATT Histo | | 2.5 mol \cdot L ⁻¹ NaCl, 100 mmol \cdot L ⁻¹ EDTA, | | | | | | 17 LC | | |
| Det (Wiston) | 母》[5] Chlornsmifee | IIIL 牟田がB D1acd ac11a | 09.0 | 0.25 mol·L ⁻¹ NaOH, | 10 | 4 | ю | 10 | 20 | V /C | 20 min | [116] |
| Kal (WISIAI) | CIII0IpyIII0S | DIOUU CEIIS | | 1% Triton X100, 10% DMSO | | | | | | | | |
| | 撒尔维亚提取物 | 结肠细胞和 | | 2.5 mol·L ⁻¹ NaCl, | | | | | | | | |
| 大鼠 | S. officinalis | 淋巴细胞 | 未提 | 100 mmol·L ⁻¹ Na ₂ EDTA, | 10 | 4 | - | > 13 | 40 | 21 V | 20 min | [117] |
| Rat (Fischer) | water extract | Colonocytes, lymphocytes | ши | 10 mmol·L ⁻¹ Tris, 1% Triton X-100 | | | | | (30 |) mA, 0.8 V•cm | - ⁻) (4 °C) | |
| | 1-氧化- | 血细胞、 | | 100 mmol·L ⁻¹ EDTA, | | | | | | | | |
| 大鼠 Rat | 4-硝基喹啉 | 肝细胞 | 29.0 | 2.5 mol \cdot L ⁻¹ NaCl, | 12 | Ţ | 过夜 | 12 | 30 | 1 V E O | 20 min | [110] |
| (Sprague-Dawley) | 4-Nitroquinoline- | Blood, | 10.0 | 10 mmol·L ⁻¹ Trizma base, | CI | + | vemight | CI | 00 | 0./ V·CIII | (4 °C) | [011] |
| | 1-oxide (4NQO) | liver cells | | 1% Triton X-100, 10% DMSO | | | | | | | | |
| | 吲哚美辛 | | | 1% Sodium sarcosinate, | | | | | | | | |
| 大鼠 | Indomethacin, | 淋巴细胞 | 01.0 | 2.5 mol·L ⁻¹ NaCl, | 0 | - | - | 5 | 00 | | | [011] |
| Rat (Wistar) | 番茄红素 | Lymphocytes | 00 | 100 mmol $\cdot L^{-1}$ Na ₂ EDTA, | 10 | t | - | <u>c1</u> | 70 | 1.0 V • CIII | 11111 07 | [411] |
| | Lycopene | | | 10 mmol·L ⁻¹ Tris, 1% Triton X-100 | | | | | | | | |

李岗等:彗星实验技术及其应用

3.3 蚯蚓彗星实验

蚯蚓的小细胞常用于彗星实验,小细胞来自蚯 蚓的体腔液,是体腔液中含量最丰富的细胞,类似于 脊椎动物的白细胞。蚯蚓体腔细胞的收集方法,包 括提取液法、穿刺法、电击法和超声波法等^[33]。

有多种蚯蚓用于彗星实验,如 Eisenia fetida^[80]、 Amynthas diffringens^[81]、Aporrectodea caliginosa^[82]、 Dendrodrilus rubidus^[81]、Lumbricus terrestris^[83]和 Microchaetus benhami^[81]等,其中 E. fetida 是最常用的 品种,为 OECD 和美国环境保护局(EPA)推荐, E. fetida 和 A. caliginosa 对污染物有相同的敏感性^[82], 而 D. rubidus 对重金属 Cd 最敏感,其次是 E. fetida^[81],例见表4。

3.4 哺乳动物精细胞彗星实验

哺乳动物的彗星实验,人和鼠最常见,涉及的细胞主要是血细胞和淋巴细胞,其他还有心、肝、肺、 肾、脑、骨髓等细胞,例见表5和表6。

常用改进的彗星实验检测精子的 DNA 损伤与 不孕的关系^[84],叫做二维垂直慧尾彗星实验 TTcomet (two-dimensional perpendicular tail comet assay, TT-comet),适合区分单链断裂和双链断裂的精细 胞^[85]。精细胞和体细胞对相同毒物的反应差别很 大,因为体细胞的异染色质是组蛋白而精细胞的则 是精蛋白^[86]。因此,去除对照的 DNA 结合蛋白的 裂解条件具有物种的特异性,不同物种的精细胞 DNA 的结构不同,因而,不同物种的 TT-comet 条件 各不相同,需要摸索。一般,去除蛋白的精细胞 DNA 先进行中性电泳,产生双链断裂的惹尾,然后 转胶 90 度,进行碱性电泳,产生单链断裂的或碱不 稳定的慧尾。

4 存在的问题与注意点(Questions and defects)

(1)大部分已经发表的文献,彗星实验设计缺少 阳性对照,降低了实验的正确性和合理性。可参考 OECD 489 推荐的彗星实验阳性对照(见表 1)。另 外,文献中提供的彗星实验图片较少,只有统计图 表,这样说服力不强,降低了文章的可靠性,在引用 时需要注意。还需要了解的是,2016 年以前的彗星 实验,以低通量为主,一次实验的处理数和重复数设 置都不是太多。

(2)彗星实验方案不统一,每种生物基本没有标准的实验方案,限制了结果的可比性。不同物种,不同细胞类型使用同一种实验,可能导致实验失败,例

如,在蚯蚓体腔细胞的彗星实验中,细胞裂解液中的 Triton-100浓度采用通常值1%,就不合适(未发表), 很难得到完美的实验图象。

(3)DNA 链的断裂损伤类型因组织和细胞类型 而异^[87]。因此,不能用血细胞 DNA 损伤类型和程 度来推断其他细胞 DNA 的损伤。如,用彗星实验 比较细鳞蝲(*Therapon jarbua*)的腮细胞、肾细胞和血 细胞对氯化汞损伤的敏感性,结果表明腮细胞最敏 感、肾细胞次之、血细胞最不敏感^[88]。同样,不能用 不同类型的细胞进行修复能力的相互推断^[60],如剑 尾鱼(*Xiphophorus species*)脑部细胞对核苷酸切除修 复能力大于腮部和肝部细胞^[89]。

(4)注意细胞周期对彗星实验结果的影响。细胞处于不同周期如 S 期或 M 期,进行彗星实验,都能进行损伤测定^[90-91],在碱性条件下 S 期的 DNA 迁移速度比中性条件下快,因为在碱性条件下 S 期的 DNA 呈复制叉状,类似单链断裂 DNA,而在中性条件下呈复制泡状^[92]。

(5)运用彗星实验结果评价毒物的 DNA 毒性时,还必须考虑非毒物相关因子,包括生物的和非生物因子对彗星实验的影响。例如,水中缺氧或氧过饱和时,都能增加鲤鱼(*Cyprinus carpio*)的 DNA 损伤,幅度约为正常氧浓度的 20%^[93]。激流中生活的白鲑(*Leuciscus cephalus*)DNA 损伤会增加,暗示生活在污染激流中的鱼面临更为严重的 DNA 损伤风险^[67]。实验生物的年龄、性别等对彗星实验结果也有影响,如成年法国比目鱼(*Limanda limanda*)面对同样的毒物,雄性 DNA 损伤比雌性高,幼鱼则相反,成年鱼比幼鱼高^[94]。此外,需要评估组织取样时使用的麻醉剂对 DNA 可能造成损伤。但是,苯唑卡因(benzocaine)对罗非鱼(*Nile tilapia*)的彗星实验结果无影响^[95]。

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通讯作者简介:蔡磊明(1962—),男,博士,教授级高级工程 师,主要研究方向为生态学、化学品安全评价及环境科学。

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